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Title: Climatic controls of decomposition drive the global biogeography of forest tree symbioses

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431 **Manuscript**

432 **The identity of the dominant microbial symbionts in a forest determines the ability**
 433 **of trees to access limiting nutrients from atmospheric or soil pools^{1,2}, sequester**
 434 **carbon^{3,4} and withstand the impacts of climate change¹⁻⁶. Characterizing the global**
 435 **distribution of symbioses, and identifying the factors that control it, are thus integral to**
 436 **understanding present and future forest ecosystem functioning. Here we generate the first**
 437 **spatially explicit global map of forest symbiotic status using a database of over 1.1 million**
 438 **forest inventory plots with over 28,000 tree species. Our analyses indicate that climatic**
 439 **variables, and in particular climatically-controlled variation in decomposition rate, are the**
 440 **primary drivers of the global distribution of major symbioses. We estimate that**
 441 **ectomycorrhizal (EM) trees, which represent only 2% of all plant species⁷, constitute**

approximately 60% of tree stems on Earth. EM symbiosis dominates forests where seasonally cold and dry climates inhibit decomposition, and are the predominant symbiosis at high latitudes and elevation. In contrast, arbuscular mycorrhizal (AM) trees dominate aseasonally warm tropical forests and occur with EM trees in temperate biomes where seasonally warm-and-wet climates enhance decomposition. Continental transitions between AM and EM dominated forests occur relatively abruptly along climate driven decomposition gradients, which is likely caused by positive plant-microbe feedbacks. Symbiotic N-fixers, which are insensitive to climatic controls on decomposition compared with mycorrhizal fungi, are most abundant in arid biomes with alkaline soils and high maximum temperatures. The climatically driven global symbiosis gradient we document represents the first spatially-explicit, quantitative understanding of microbial symbioses at the global scale and demonstrates the critical role of microbial mutualisms in shaping the distribution of plant species.

Microbial symbionts strongly influence the functioning of forest ecosystems. They exploit inorganic, organic² and/or atmospheric forms of nutrients that enable plant growth¹, determine how trees respond to elevated CO₂⁶, regulate the respiratory activity of soil microbes^{3,8}, and affect plant species diversity by altering the strength of conspecific negative density dependence⁹. Despite growing recognition of the importance of root symbioses for forest functioning^{1,6,10} and the potential to integrate symbiotic status into Earth system models that predict functional changes to the terrestrial biosphere¹⁰, we lack spatially-explicit, quantitative maps of the different root symbioses at the global scale. Generating these quantitative maps of tree symbiotic states would link the biogeography of functional traits of belowground microbial

symbionts with their 3.1 trillion host trees¹¹, spread across Earth's forests, woodlands, and savannas.

The dominant guilds of tree root symbionts, arbuscular mycorrhizal (AM) fungi, ectomycorrhizal (EM) fungi, ericoid mycorrhizal (ErM) fungi, and nitrogen (N)-fixing bacteria (N-fixer) are all based on the exchange of plant photosynthate for limiting macronutrients. The AM symbiosis evolved nearly 500 million years ago, with EM, ErM and N-fixer plant taxa evolving multiple times from an AM basal state. Plants that form the AM symbiosis comprise nearly 80% of all terrestrial plant species, and principally rely on AM fungi for enhancing mineral phosphorus (P) uptake¹². In contrast to AM fungi, EM fungi evolved from multiple lineages of saprotrophic ancestors, and as a result some EM fungi are more capable of mobilizing organic sources of soil nutrients (particularly nitrogen)². Association with EM fungi, but not AM fungi, has been shown to allow trees to accelerate photosynthesis in response to increased atmospheric CO₂ when soil nitrogen (N) is limiting⁶ and to inhibit soil respiration by decomposer microbes^{3,8}. Because increased plant photosynthesis and decreased soil respiration both reduce atmospheric CO₂ concentrations, the EM symbiosis is associated with buffering the Earth's climate against anthropogenic changes.

In contrast to mycorrhizal fungi, which extract nutrients from the soil, symbiotic N-fixers (Rhizobia and Actinobacteria) convert atmospheric N₂ to plant-usable forms. Symbiotic N-fixers are responsible for a large fraction of biological soil-N inputs, which can increase N-availability in forests where they are locally abundant¹³. Both N-fixing bacteria and EM fungi often demand more plant photosynthate than does the AM symbiosis^{12,14,15}. Because tree growth and reproduction are limited by access to inorganic, organic and atmospheric sources of N, the distribution of root symbioses is likely to reflect both environmental conditions that maximize

the cost-benefit ratio of symbiotic exchange as well as physiological constraints on different symbionts.

In one of the earliest efforts to understand the functional biogeography of plant root symbioses, Sir David Read¹⁶ categorically classified biomes by their perceived dominant mycorrhizal type and hypothesized that seasonal climates favor hosts associating with EM fungi due to their ability to compete directly for organic N. In contrast, it has been proposed that sensitivity to low temperatures has prevented N-fixers from dominating outside the tropics, despite the potential for N-fixation to alleviate N-limitation in boreal forests^{15,17}. However, global scale tests of these proposed biogeographic patterns and their climate drivers are lacking. To address this research gap, we compiled the first global ground-sourced survey database to reveal numerical abundances of each symbiosis across the globe, which is essential for identifying the potential mechanisms underlying transitions in forest symbiotic state along climatic gradients^{18,19}.

We determined the abundance of tree symbioses using GFBi, an extension from the plot-based Global Forest Biodiversity (GFB) database, which contains over 1.1 million forest inventory plots of individual-based measurement records from which we derive abundance information for entire tree communities (Figure 1). Using published literature on the evolutionary histories of mycorrhizal and N-fixer symbioses, we assigned plant species from the GFBi to one of 5 symbiotic guilds: AM, EM, ErM, N-fixer, and non- or weakly-mycorrhizal (NM). We then used the random forest algorithm with K-fold cross validation to determine the importance and influence of variables related to climate, soil chemistry, vegetation, and topography on the relative abundance of each tree-symbiotic guild (Figure 2). Because decomposition is the dominant process by which soil nutrients become available to plants, we

calculated annual and quarterly decomposition coefficients according to the Yasso07 model²⁰, which describes how temperature and precipitation gradients influence mass-loss rates of different chemical pools of leaf litter (with parameters fit using a previous global study of leaf decomposition, Figures 3, S5). Finally, we projected our predictive models across the globe over the extent global biomes that fell within the multivariate distribution of our model training data (Figures 4, S14-15, see Methods for full description).

Our analysis shows that the three most numerically abundant tree symbiotic guilds each have reliable environmental signatures, with the four most important predictors accounting for 81, 79, and 52% of the total variability in EM, AM, and N-fixer relative basal area, respectively. Models for ErM and NM lack strong predictive power given the relative rarity of these symbiotic states amongst trees, although the raw data do identify some local abundance hotspots for ErM (Figure S1). As a result, we focus the remainder of results and discussion on the three major tree symbiotic states (EM, AM, N-fixer). Despite the fact that data from N. and S. America constitute 65% of the training data (at the 1 by 1 degree grid scale), our models accurately predict the proportional abundances of the three major symbioses across all major geographic regions (Figure S10). The high performance of our models, which is robust to both K-fold cross-validation and rarefying samples so that all continents are represented with equal depth (Figures S11-12), suggest that regional variations in climate (including indirect effects on decomposition) and soil pH (for N-fixers) are the primary factors influencing the relative dominance of each guild at the global scale (geographic origin only explained ~2-5% of the variability in residual relative abundance) (Table S8, Figure S10).

Whereas a recent global analysis of root traits concluded that plant evolution has favored reduced dependence on mycorrhizal fungi²¹, we find that trees associating with the relatively

more C-demanding and recently-derived EM fungi^{12,14} represent the dominant tree-symbiosis. By taking the average proportion of EM trees, weighted by spatially-explicit global predictions for tree stem density¹¹, we estimate that approximately 60% of trees on earth are EM, despite the fact that only 2% of plant species associate with EM fungi (vs. 80% associating with AM fungi)⁷. Outside of the tropics, the estimate for EM relative abundance increases to approximately 80% of trees.

Turnover among the major symbiotic guilds results in a tri-modal latitudinal abundance gradient, with the proportion of EM trees increasing (and AM trees decreasing) with distance from the equator, while the upper-quantiles of N-fixing trees reach peak abundance in the arid zone around 30 degrees (Figure 3A, Figure 4). These trends are driven by abrupt transitional regions along continental climatic gradients (Figure 2), which skew the distribution of symbioses among biomes (Figure 3A) and drive strong patterns across geographic and topographic features that influence climate. Moving north or south from the equator, the first transitional zone separates warm (aseasonal), AM-dominated, tropical broadleaf forests (>75% median basal area, vs. 8% for EM trees) from the rest of the EM-dominated world forest system (Figure 2AB; Figure 3A). The transition zone occurs across the globe around 25 degrees N and S latitude, just beyond the dry tropical broadleaf forests (with 25% EM tree basal area; Figure 3A), where average monthly temperature variation reaches 3-5°C (temperature seasonality, Figure 2AB).

Moving further N or S, the second transitional climate zone separates regions where decomposition coefficients during the warmest quarter of the year are less than 2 (see Figure 3B for the associated temperature and precipitation ranges). In N. America and China, this transition zone occurs around 50 degrees N, separating the mixed AM / EM temperate forests from their neighboring EM dominated boreal forests (75 vs 100% EM tree basal area, respectively; Figure

3A). This transitional decomposition zone bypasses W. Europe, which has temperature seasonality $> 5^{\circ}\text{C}$, but lacks sufficiently wet summers to accelerate decomposition coefficients beyond values associated with mixed AM/EM forests. The latitudinal transitions in symbiotic state observed among biomes are mirrored by within-biome transitions along elevation gradients. For example, in tropical Mexico, warm and wet quarter decomposition coefficients < 2 occur along the slopes of the Sierra Madre, where mixed AM-exclusive and N-fixer woodlands in arid climates transition to EM dominated tropical coniferous forests (75% basal area, Figure 3A, Figure 4ABC, Figure S16-18). The southern hemisphere, which lacks the landmass to support extensive boreal forests, experiences a similar latitudinal transition in decomposition rates along the ecotone separating its tropical and temperate biomes, around 28 degrees S.

The abrupt transitions that we detected between forest symbiotic states along environmental gradients suggest that positive feedbacks may exist between climatic and biological controls of decomposition^{10,20}. In contrast to AM fungi, some EM fungi can use oxidative enzymes to mineralize organic nutrients from leaf litter, converting nutrients to plant-usable forms^{2,5}. Relative to AM trees, the leaf litter of EM trees is also chemically more resistant to decomposition, with higher C:N ratios and higher concentrations of decomposition-inhibiting secondary compounds¹⁰. Thus, EM leaf litter can exacerbate climatic barriers to decomposition, promoting conditions where EM fungi have superior nutrient-acquiring abilities to AM-fungi^{5,10}. A recent game theoretical model has shown that positive plant-soil-nutrient feedbacks can lead to local bistability in mycorrhizal symbiosis²². Such positive-feedbacks are also known to cause abrupt ecosystem transitions along smooth environmental gradients between woodlands and grasses: trees suppress fires, which promotes seedling recruitment, while grass fuels fires, which kill tree seedlings²³. The existence of abrupt transitions also suggests that forests in transitional

regions along decomposition gradients should be susceptible to drastic turnover in symbiotic state with future environmental changes²³.

To illustrate the sensitivity of global patterns of tree symbiosis to climate change, we use the relationships we developed for current climate to project potential changes in forest symbiotic status in the future. Relative to our global predictions using the most recent climate data, model predictions using the projected climates for 2070 suggest the abundance of EM trees will decline by as much as 10% (using a relative concentration pathway of 8.5 W/m²; Figure S24). Due to their position along decomposition gradients relative to the abrupt shift from EM to AM forests (Figure 2AB), our models predict the largest declines in EM abundance will occur along the boreal-temperate ecotone, although this model does not estimate the time lags between climate change and forest community responses. The predicted decline in EM trees corroborates the results of common garden transfer and simulated warming experiments, which demonstrate that some important EM hosts will decline at the boreal-temperate ecotone in altered climates²⁴.

The change in dominant nutrient exchange symbioses along climate gradients highlights the interconnection between atmospheric and soil compartments of the biosphere. The transition from AM to EM dominance corresponds with a shift from P to N limitation of plant growth with increasing latitude^{25,26}. Including published global projections of total soil N or P, microbial N, or soil P fractions (labile, occluded, organic, and apatite) did not increase the amount of variation explained by the model or alter the variables identified as most important, and thus were dropped from our analysis. However, our finding that climatic controls of decomposition best predict the dominant mycorrhizal associations mechanistically links symbiont physiology with climatic controls of soil nutrient release from leaf litter. These findings are consistent with Read's hypothesis¹⁶ that slow decomposition at high latitudes favors EM fungi due to their increased

capacity to liberate organic nutrients². Thus, while more experiments are necessary to understand the specific mechanism by which nutrient competition favors dominance of AM or EM symbioses¹⁸, we propose that the latitudinal and elevational transitions from AM to EM dominated forests be called Read's Rule.

While our analyses focus on prediction at large spatial scales appropriate to the available data, our findings with respect to Read's Rule also provide insight into how soil factors structure the fine-scale distributions of tree symbioses within our grid cells. For example, while at a coarse scale we find that EM trees are relatively rare in many wet tropical forests, individual tropical sites in our raw data span the full range from 0 – 100 % EM basal area. In much of the wet tropics, these EM dominated sites exist as outliers within a matrix of predominantly AM trees. In an apparent exception that proves Read's Rule, in aseasonal warm neotropical climates, which accelerate leaf-decomposition and promote regional AM dominance (Figure 3), EM dominated tree stands can develop in sites where poor soils and recalcitrant litter slow decomposition and N mineralization^{18,27}. Landscape-scale variation in the relative abundance of symbiotic states also changes along climate gradients, with variability highest in xeric and temperate biomes (Figure S3-4), suggesting that the potential of local nutrient variability to favor particular symbioses is contingent on climate.

Whereas EM trees are associated with ecosystems where plant growth is thought to be primarily N-limited, N-fixer trees are not. Our results highlight the global extent of the “N-cycling paradox,” wherein some metrics suggest that N-limitation is greater in the temperate zone^{25,26}, yet N-fixing trees are relatively more common in the tropics^{15,28} (Figure 3A). We find that N-fixers, which we estimate represent 7% of all trees, dominate forests with annual max temperatures >35°C and alkaline soils (particularly in North America and Africa, Figure 2C).

They have the highest relative abundance in xeric shrublands (24%), tropical savannas (21%), and dry broadleaf forest biomes (20%), but are nearly absent from boreal forests (<1%) (Figure 3A, Figure 4). The decline in N-fixer tree abundance we observed with increasing latitude is also associated with a previously documented latitudinal shift in the identity of N-fixing microbes, from facultative N-fixing rhizobial bacteria in tropical forests to obligate N-fixing actinorhizal bacteria in temperate forests²⁸. Our data are not capable of fully disentangling the several hypotheses that have been proposed to reconcile the N-cycling paradox¹⁵. However, our results are consistent with the model prediction¹⁷ and regional empirical evidence^{19,29,30} that N-fixing trees are particularly important in arid biomes. Based primarily on the observed positive, nonlinear association of N-fixer relative abundance with the mean temperature of the hottest month (Figure 2C), our models predict a two-fold increase in N-fixer relative abundance when transitioning from humid to dry tropical forest biomes (Figure 3A).

Although soil microbes are a dominant component of forests, both in terms of diversity and ecosystem functioning^{5,6,10}, identifying global-scale microbial biogeographic patterns remains an ongoing research priority. Our analyses confirm that Read's Rule, which is one of the first proposed biogeographic rules specific to microbial symbioses, successfully describes global transitions between mycorrhizal guilds. More generally, climate driven turnover among the major plant-microbe symbioses represents a fundamental biological pattern in the Earth system, as forests transition from low-latitude arbuscular mycorrhizal, to N-fixer, to high-latitude ectomycorrhizal ecosystems. The predictions of our model (which we make available as a global raster layer) can now be used to represent these critical ecosystem variations in global biogeochemical models used to predict climate-biogeochemical feedbacks within and between trees, soils, and the atmosphere. Additionally, the layer containing the proportion of N-fixing

trees can be used to map potential symbiotic N-fixation, which links together atmospheric pools of C and N. Future work can extend our findings to incorporate multiple plant growth forms and non-forested biomes, where similar patterns likely exist, to generate a complete global perspective. Our predictive maps leverage the most comprehensive global forest dataset to generate the first quantitative global map of forest tree symbioses, demonstrating how nutritional mutualisms are coupled with the global distribution of plant communities.

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Figure Legends

Figure 1. The global distribution of GFBi training data. The global map has n=2,768 grid cells at a 1 x 1 degree latitude/longitude resolution. Cells are colored in the red, green and blue spectrum according to the % of total tree basal area occupied by N-fixer, AM, and EM tree symbiotic guilds, as indicated by the ternary plot. Grey cells show the global distribution of forests where we make model projections.

Figure 2. A small number of environmental variables predict the majority of global turnover in forest symbiotic status. Panels show the partial feature contributions of different environmental variables on forest symbiotic state. Each row plots the shape of the contribution of the four most important predictors of the proportion of tree basal area belonging to the (a) ectomycorrhizal (EM), (b) arbuscular mycorrhizal (AM), and (c) N-fixer symbiotic guilds (n=2,768). Variables are listed in declining importance from left to right, as determined by inc node purity, with points colored with a red-green-blue gradient according to their position on the x-axis of the most important variable (left-most panels for each guild), allowing cross visualization between predictors. Each panel lists two measures of variable importance, inc node purity (used for sorting) and %IncMSE (see Supplemental Information for description). The abundance of each symbiont type transitions sharply along climatic gradients, suggesting that sites near the threshold are particularly vulnerable to switching their dominant symbiont guild with climate changes.

Figure 3. The distribution of forest symbiotic status across biomes is related to climatic controls over decomposition. (a) Biome level summaries of the median +/- 1 quartile of the predicted % tree basal area per biome for ectomycorrhizal (EM), arbuscular mycorrhizal

(AM), and N-fixer symbiotic guilds (n=100 random samples per biome). (b) The dependency of decomposition coefficients (k, solid and dotted lines) on temperature and precipitation during the warmest quarter with respect to predicted dominance of mycorrhizal symbiosis. The transition from AM forests to EM forests between k=1 and 2 is abrupt, which is consistent with positive feedback between climatic and biological controls of decomposition.

Figure 4. Global maps of predicted forest tree symbiotic state. Maps (left) and latitudinal gradients (right, with solid line indicating the median and colored ribbon spanning the range from the 5% and 95% quantiles) of the % of tree basal area for (a) ectomycorrhizal (EM), (b) arbuscular mycorrhizal (AM), and (c) N-fixer symbiotic guilds. All projections are displayed a 0.5 by 0.5 degree lat/long scale with n=28,454.

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Author Contributions

KGP & TWC conceived the study; TWC, JL, PBR, GN, SdM, MZ, NP, BH, XZ, & CZ conceived and organized the GFBi database; KGP, BSS, GDAW, & MVN compiled the symbiosis database; BSS carried out the primary data analysis; MVN & DR contributed to data compilation and analysis; BSS, TWC, MVN & KGP wrote the initial manuscript; BSS, TWC, JL, MVN, GDAW, PBR, GN, SdM, MZ, NP, BH, XZ, CZ & KGP made substantial revisions to

all versions of the manuscript; all other named authors provided forest inventory data and commented on the manuscript.

Data Availability

The GFBi database is available upon written request at <https://www.gfbinitiative.org/datarequest>. Additionally, the symbiotic state assigned to tree species as a supplementary file, as are global rasters of our model projections for EM, AM, and N-fixer proportion of tree basal area.

Conflict of Interest

The authors declare we have no conflict of interests.

Supplementary Information

For more information on symbiotic guild assignments, model selection, and supplementary analyses, refer to SupplementaryInfo_Steidinger_etal2019.pdf. For the full suite of Supplemental Files, including symbiotic guild assignments and rasters of model projections, refer to SupplementalFiles_Steidinger_etal2019.zip

Methods

We quantified the relative abundance of tree symbiotic guilds across >1.1 million forest census plots combined in the GFBi database, an extension from the plot-based Global Forest Biodiversity (GFB³¹) database. The GFBi database consists of individual-based data that we compiled from all the regional and national GFBi forest inventory data sets. The standardized GFBi data frame, i.e. tree list, comprises tree ID, a unique number assigned to each individual tree; plot ID, a unique string assigned to each plot; plot coordinates, in decimal degrees of WGS84 datum; tree size, in diameter-at-breast-height; trees-per-hectare expansion factor; year of

measurement; data set name, a unique number assigned to each forest inventory data set; and binomial scientific tree species names.

We error checked all species names from different forest inventory data sets in three steps. First, we extracted scientific names from original data sets, keeping only the names of genus and species (authority names are removed). Next, we compiled all the species names into five general species lists, one for each continent. Finally, we verified individual species names against 23 online taxonomic databases using the ‘taxize’ package of R programming language³². We assigned each morphospecies a unique name comprising the genus, the string “spp”, followed by the data set name and a unique number for that species. For example, “Picea sppCNI1” and “Picea sppCNI2” represent two different species under the genus “Picea”, observed in the first Chinese data set (CNI).

We derived plot-level abundance information in terms of species abundance matrices. Each species abundance matrix consisted of the number of individuals by species (column vectors) within individual sample plots (row vectors). In addition, key plot-level information was also added to the matrices, including plot ID, data set name, plot coordinates, the year of measurement, and basal area, i.e. the total cross-sectional areas (m²) of living trees per one hectare of ground area.

Tree genera were assigned to a plant family using a plant taxonomy lookup table generated by Will Cornwell (hosted on Github <https://github.com/traitecoevo/taxonlookup>), which uses the accepted taxonomy from “The Plant List.” The majority (96.5%) of genera from the GFBi species were successfully matched to family; for those that could not be assigned, we manually checked the GFBi genus and species against synonyms from The Plant List. Of the remaining 1,038 mismatches, an additional 440 were assigned to family either by updating older

genera and species names with their more recent synonyms or else by correcting obvious misspellings. The remaining 598 entries that could not be matched to family were excluded from analysis.

We used a taxonomically-informed approach to assign symbiotic states to plant species from the GFBi. Plant species were assigned to one of 5 symbiotic guilds – ectomycorrhizal (EM), arbuscular mycorrhizal (AM), ericoid mycorrhizal (ErM), weakly AM or non-mycorrhizal (AMNM), or N-fixer (Table S1). Although we did not model the relative abundance of ErM trees, due to their rarity, we have included a map of their relative abundance from our grid (Figure S1). We also include as a supplementary file the full species list, which includes columns used to assign species to guild. In addition, we include here a list of families and genera assigned to all guilds except AM (Tables S2-5) with notes for cases of species from individual genera that were either assigned to two guilds simultaneously (e.g., *Alnus* is an N-fixer and EM) or where species from individual genera were split between two different guilds (e.g., some *Pisonia* sp. are weakly AM and some are EM). An AM summary table is excluded for length considerations—the same information is available in the Supplementary File “SymbioticGuildAssignment.csv”.

The taxonomy of species in our inventory was compared with recently published literature on the evolutionary history of mycorrhizal symbiosis^{7,33,34} and N-fixation³⁵⁻³⁸. Most species symbiotic status could be reliably assigned at the genus (e.g. *Dicymbe*) or family level (e.g. Pinaceae). For the few groups where status was unreliable or variable within a genus (e.g. *Pisonia*) we conducted additional literature searches.

We assigned species to the EM category in three stages. First, at the family level (e.g., Pinaceae); next, at the genus level (e.g., *Dicymbe*); and finally, using literature searches for unclear genera. For example, for the genus *Pisonia*, some species are AM and others are EM. We

used a published list from Hayward & Hynson (2014)³⁹ to sort species into the appropriate guild. For the genus *Acacia*, we followed Brundrett (2017)⁷ in assuming that only endemic Australian species associate with EM, while all others are AM (we sorted *Acacia* species according to provenance using <http://worldwidewattle.com/>).

The AMNM category lumped together all genera of terrestrial, non-epiphytic plants that either lack arbuscular mycorrhizal fungi (AMF), or have low or inconsistent records of AMF colonization of roots. For example, although there are some published records of AMF colonization in the roots of Proteaceae, these records are inconsistent, and colonization is generally low. Further, as Proteaceae are associated with a non-mycorrhizal root morphology (the “cluster” or “proteoid root system”) that allows them to access otherwise unavailable forms of soil nutrients⁴⁰, we placed the entire family within AMNM. The family Urticaceae, which we also characterized as AMNM, was somewhat problematic – early-successional species from tropical forests, such as those in the genus *Cecropia*, have records of both low and absent AMF colonization⁴¹. Our approach was to use the most broadly inclusive AMNM categorization.

N-fixer status was assigned at the genus level, using previously compiled databases of global symbiotic N₂-fixation³⁵⁻³⁸. Given that symbiotic N₂-fixation with rhizobial or *Frankia* bacteria has only evolved in four orders (Rosales, Cucurbitales, Fabales and Fagales)⁴², all species outside of this nitrogen-fixing clade were assigned non-fixing status. Some species could not be assigned a N-fixer status because they were typed to a higher taxonomic level (e.g. family) that is ambiguous from a N-fixer status perspective. We recorded when our assignment of N-fixer status was based on phylogenetic criteria but where symbiotic N-fixation is evolutionarily labile. Since these cases are more likely to be misassigned we excluded them from the N-fixation category. The N-fixer group contains species that are colonized by AMF (e.g.,

most genera from Leguminosae) and others that are colonized by ectomycorrhizal fungi (e.g., *Alnus* sp.).

Most plant species form AM symbiosis, which is the basal symbiotic state to the later derived EM and N-fixing symbioses. Further, many EM and N-fixing plants maintain the ability to form AM symbiosis. Thus, a tree species is most likely AM if it does not form associations with another symbiotic guild (or forgoes root symbiosis entirely), as evidenced by their inclusion in exhaustive databases of plant symbiotic state^{7,33-38,41}. In keeping with other large-scale studies in the field (e.g. ³⁴), we assigned tree species from the GFBi database to an AM-exclusive state if they belonged to taxa that were not matched to EM, ErM, non-or-weakly mycorrhizal or N-fixer symbioses. Thus, the AM and N-fixer groups in our dataset are non-overlapping despite the fact that most N-fixers also associate with AM fungi.

The proportions of tree basal area and tree individuals were aggregated to a 1' by 1' degree grid by taking the weighted average of the plot-level proportions (Table S6). This resulted in a total of 2,768 grid cells, each with a score for the proportional abundance of EM, AM, N-fixer, ErM, and AMNM trees. We calculated two measures of relative abundance for each symbiotic guild: proportion of tree stems and proportion of tree basal area. Because the measurements are highly correlated with one another (Figure S2) we chose to model only proportion of total tree basal area, which should scale more approximately to proportion of tree biomass as it accounts for differences in size among individual stems. Additionally, we quantified variability among plots within each grid cell by calculating the weighted standard deviation across the grid (Supplemental Information, Figure S3-4).

To identify the key factors structuring symbiotic distributions we assembled 70 global predictor layers: 19 climatic (annual, monthly, and quarterly temperature and precipitation

variables), 14 soil chemical (total soil N density, microbial N, C:N ratios and soil P fractions, pH, cation exchange capacity), 5 soil physical (soil texture and bulk density), 26 vegetative indices (leaf area index, total stem density, enhanced vegetation index means and variances), and 5 topographic variables (elevation, hillshade) (Table S7). Because decomposition is the dominant process by which soil nutrients become available to plants, we generated 5 additional layers that estimate the climatic control of decomposition. We parameterized decomposition coefficients according to the Yasso07 model^{20,43} using the following equation:

$$k = \text{Exp}(0.095T_i - 0.00014 T_i^2) (1 - \text{Exp}[-1.21 P_i]), \quad (1)$$

where P_i and T_i are precipitation and mean temperature, either quarterly or annually, and the constants 0.0095 ($=\beta_1$) \pm 0.00014 ($=\beta_2$), and -1.21 ($=\gamma$) are parameters fit using a previous global study of leaf litter mass-loss²⁰. Although local decomposition rates can vary significantly based on litter quality or microbial community composition⁴⁴, climate is the primary control at the global scale²⁰. Decomposition coefficients describe how fast different chemical pools of leaf litter lose mass over time relative to a parameter, α , that accounts for leaf-chemistry. Decomposition coefficients (k) with values of 0.5 and 2 indicate a halving and doubling of decomposition rates relative to α , respectively (Supplemental Information, Figure S5).

We implemented the random forest algorithm using the “randomForest” packaged in R. Random forest models average over multiple regression trees, each of which uses a random subset of all the model variables to predict a response. We first determined the influence and relationship of all 75 predictor layers on forest symbiotic state and then optimized our models using a stepwise reduction in variables, from least- to most-important. Variable importance was measured in two ways: Inc Node Purity and %IncMSE (with values reported in each panel of Figure 2). The inc node purity of variable x considers the decrease in the residual sum of squares

that results from splitting regression trees using variable x . %IncMSE (mean square error) quantifies the increase in model error as a result of randomly shuffling the order of values in the vector x . We chose to rank variables according to inc node purity because we found that higher inc node purities were associated with larger effect sizes, whereas larger %IncMSE were associated with more linear responses of smaller effect. Whereas our inspection of partial feature contributions is derived from univariate random forest models, we additionally ran multivariate random forests to predict the proportional abundance of EM, AM, and N-fixer trees for each pixel. The multivariate models were run using 50-regression trees each, with the unique set of the best 4 predictor variables for each symbiotic guild in the univariate models (Table S7, Figure 2). Despite strong negative correlations between the proportions of EM and AM basal area (Figure S22), the results from multivariate and univariate random forests are strongly correlated with one another (Figure S23).

Using model selection based on eliminating variables with low Inc Node Purity, we removed most soil nutrient, vegetative, and topographic variables from our models (Figure S6-7). Our final models include the remaining 34 predictor layers with climate, decomposition, and certain soil physical and chemical information (Figure S8). To determine the parsimony of our models, we compared the coefficient of determination in models run with a stepwise reduction in the number of variables (starting with those with the lowest Inc Node Purity). Based on performance of the ratio of coefficient of determination in models with 4 vs 34 variables, we determined that the 4 most important variables accounted for >85% of the explained variability (Figure S9). We also compared model performance visually with plots of actual vs predicted proportions of each tree symbiotic guild among continents and geographic subregions (Figure S10). We used the “forestFloor” packaged in R to plot the partial variable response of tree

935 symbiotic guilds to each predictor variable (Figure 2ABC, see Figure S19-21 for partial plots of
936 the partial feature contributions of all 34 variables).

937 In order to test the sensitivity of model performance and predictions, we performed cross
938 validation in R using the “rfUtilities” package²⁴. K-fold cross validation tests the sensitivity of
939 model predictions to losing random subsets from the training data. For EM, AM, and N-fixer
940 models we ran 99 iterations that withheld 10% of the model training data. We assessed the drop
941 in model performance in the 99 iterations by manually calculating the coefficient of
942 determination, which uses the following formula: $1 - \frac{\sum (\text{actual \% basal area} - \text{predicted \% basal area})^2}{\sum (\text{actual \% basal area} - \text{mean actual \% basal area})^2}$. For all symbiotic guilds,
943 withholding 10% of the training data resulted in a mean loss in variance explained of less than
944 1% (Figure S11). This shows that our training data has sufficient redundancy to ensure that our
945 model conclusions are robust. Similarly, to determine whether our random forest models would
946 make similar predictions if data were equally distributed among continents, we rarefied our
947 aggregated grid of symbiotic states and predictor layers to an even depth. Specifically, we sub-
948 sampled all continents – N. America (including Central America and the Caribbean), S. America,
949 Europe, Asia, and Oceania – to match the number of grid-pixels from Africa (n=50). This is a
950 much more aggressive reduction of training data than is typically used in K-fold cross
951 validations, as it involves dropping ~90% of training data rather than retaining the same amount.
952 We performed 99 iterations of rarefaction each for the three symbiotic guilds. On average,
953 models run with the rarefied data explained about 10% less variance over the full training data
954 (the entire predictor / response grid) than did models run with all of the training data (Figure
955 S12-13).

To avoid projecting our random forest models outside the ranges of their training data (e.g., grid cells with higher mean annual temperatures than the maximum used to fit the models), we subset a global grid of predictor layers depending on whether (1) the grid cell fell within the top 60% of land surface with respect to tree stem density¹¹ and either (2) fell within the univariate distribution of all the predictor layers from our training data and/or (3) fell within an 8-dimensional hypervolume defined by the unique set of the 4-best predictors of the relative abundance of each guild (Figure S14). We then projected our models across only those grid cells that met these criteria, which constitutes 46% of the global land surface and 88% of global tree stems (Figure 1; Figure S15). Model projections were made at two resolutions: both 1 by 1 degree and 0.5 by 0.5 degree resolution (Figure 4). While model validation indicates that our projections are robust, additional ground truthing of predictions to identify any discrepancies would be incredibly valuable. If such discrepancies exist they can help fine tune climate-symbiosis models, or identify areas where climate might favour invasion by symbioses that have not yet evolved or dispersed to a particular biogeographic region.

We used the following equation to estimate the % of global tree stems that belong to each tree symbiotic guild: $\sum_i (\text{predicted proportion of trees of guild } g \text{ in pixel } i) \times (\text{total number of tree stems in pixel } i) / \sum_i (\text{total number of tree stems in pixel } i)$. The proportion of tree stems and the proportion of tree basal area in each guild are highly correlated throughout the training data (Figure S4). The figures cited in the main text for each guild were calculated using model projections across all pixels, even those that did not meet the criteria for model projection because they fell outside the multivariate distribution of the predictor layers or had insufficient stem density. However, our estimates for the global % of trees occupied by each tree symbiotic guild change by <1% when using only those pixels that met our criteria for model projection.

In the main manuscript we state that sharp transitions between dominant symbiotic states with climate variables could lead to declines in EM trees, particularly in southern boreal forests. To determine this, we projected our random forest models for each symbiotic guild using climate change projections over our 19 bioclimatic variables (Table S7), including the decomposition coefficients that use temperature and precipitation values. Specifically, we considered the 2070 scenario with a relative concentration pathway (RCP) of 8.5 (W/m²), which predicts an increase of greenhouse gas emissions throughout the 21st century⁴⁵. We plot difference in the proportion of forest basal area between the projections for 2070 and those using current climate data (Table S7, Figure S24). We qualify this prediction with the note that vegetative changes to forests are constrained by rates of mortality, recruitment, and growth.

After training and cross-validating our models with GFBi data exclusively, we additionally tested whether our models accurately predicted the symbiotic state of Eurasian forests previously published by Schepaschenko et al. (2017)⁴⁶. We assigned symbiotic status to all trees in Schepaschenko et al. (2017) and aggregated plot level data to a 1 by 1 degree grid using the same methods as with the GFBi dataset (Figure S25). We found that, on average, our models predicted the symbiotic state in the regional dataset within 13.6% of the value of this other dataset (Figure S26). For projected maps in Figure 4abc, we included the Schepaschenko et al. (2017) data with the GFBi training data to increase geographic coverage throughout Eurasia.

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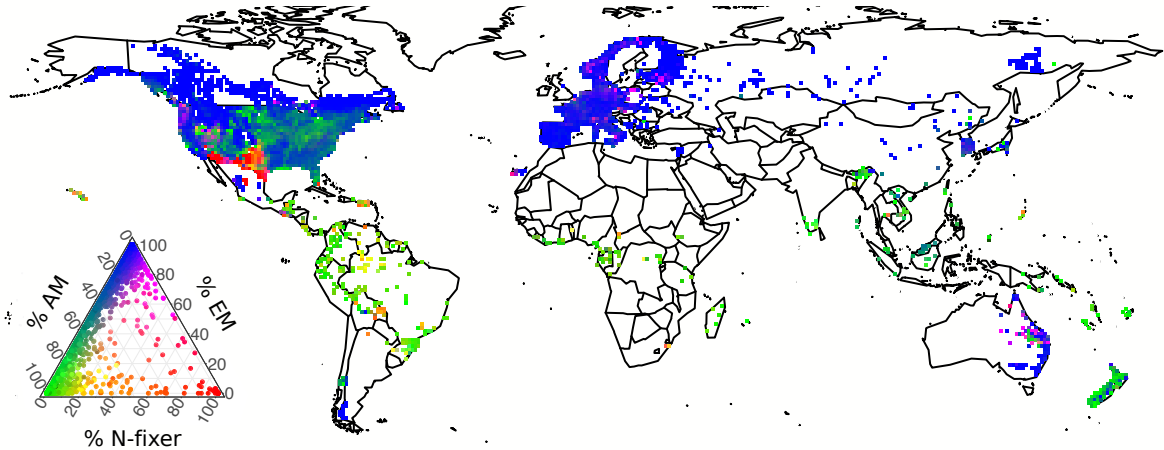
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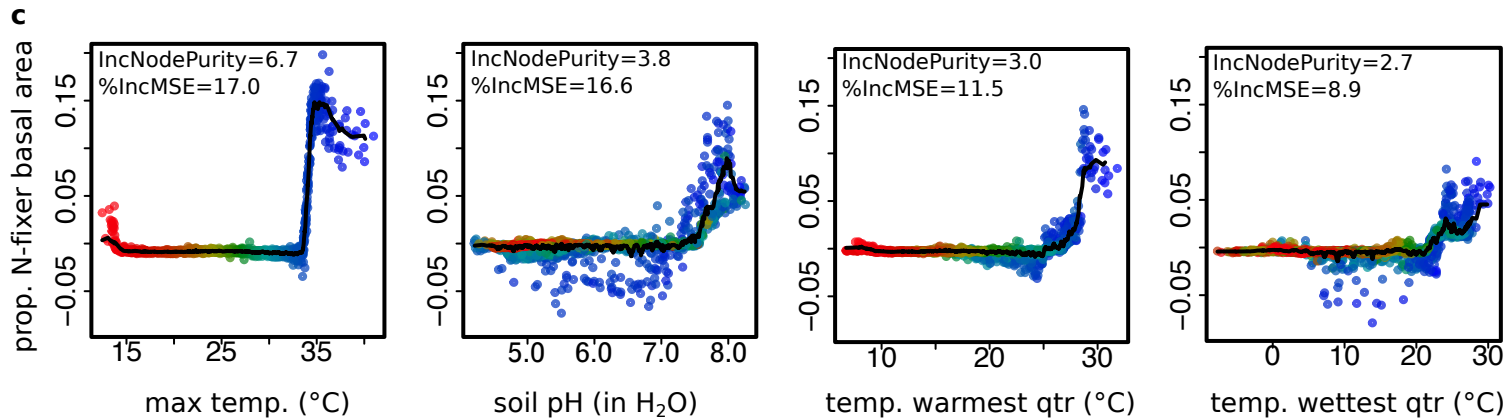
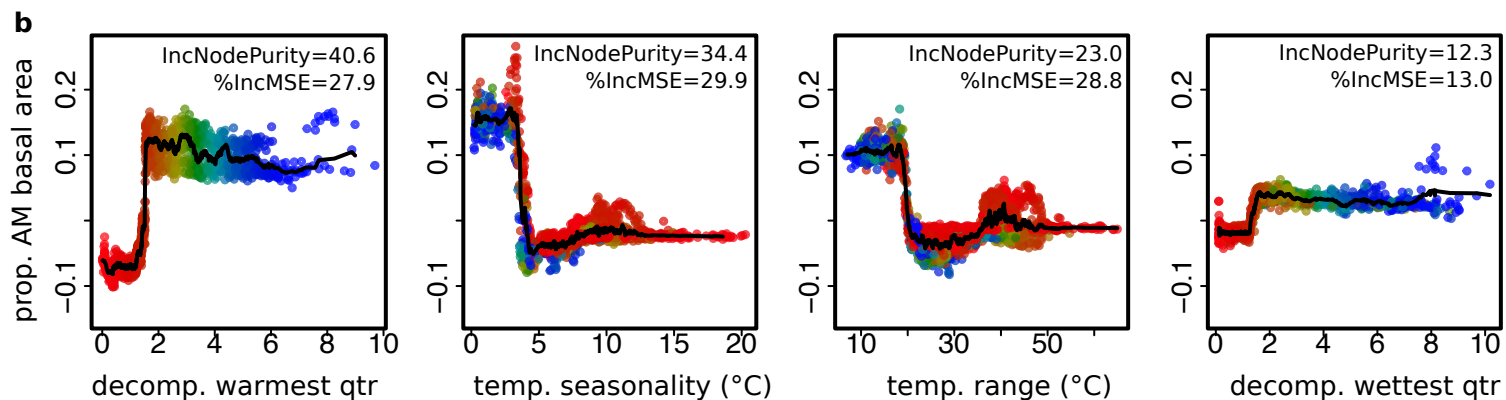
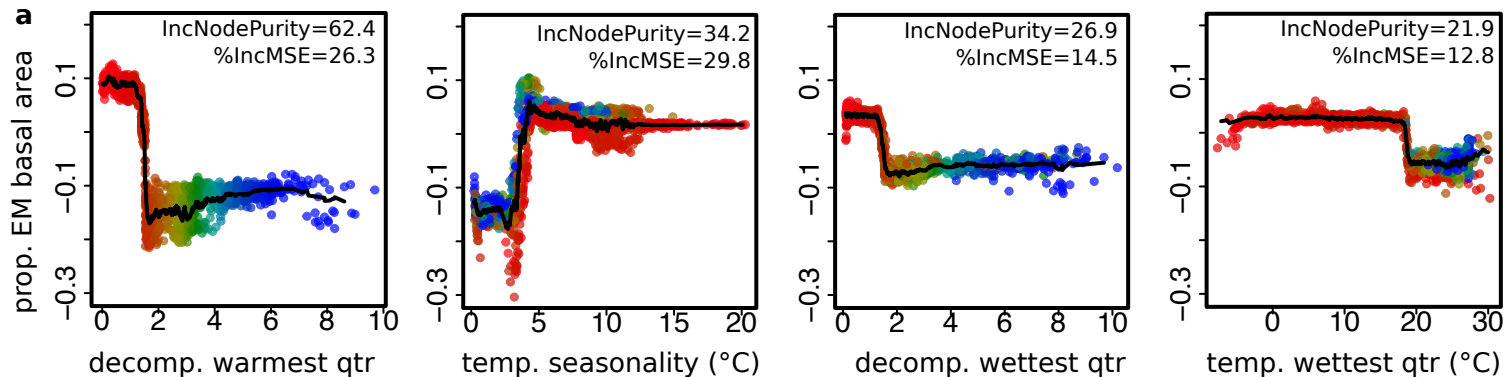
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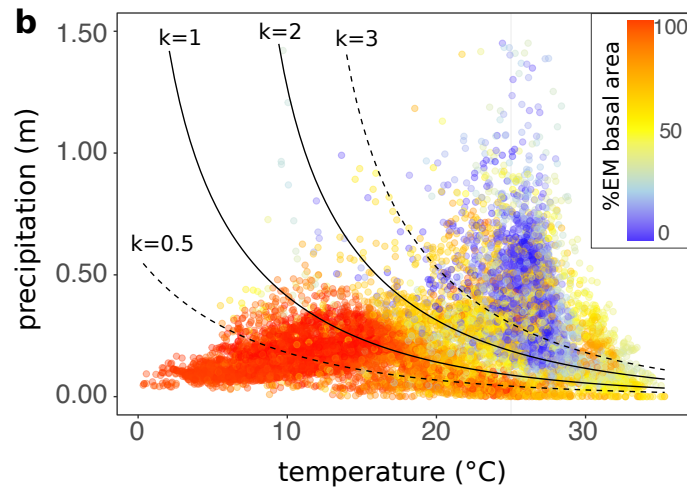
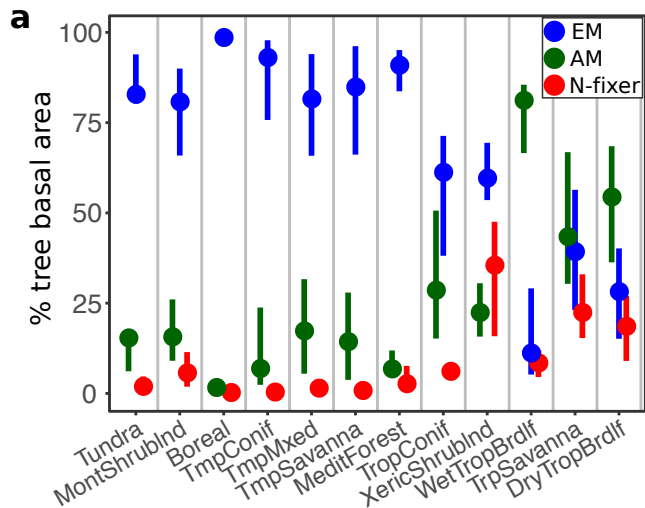
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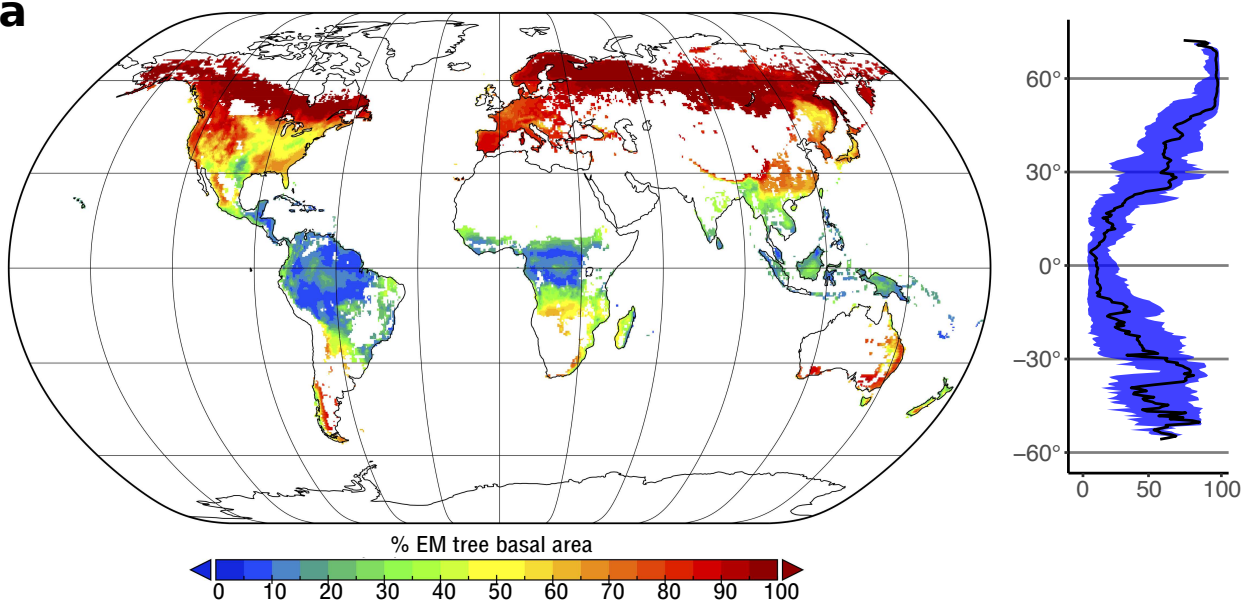
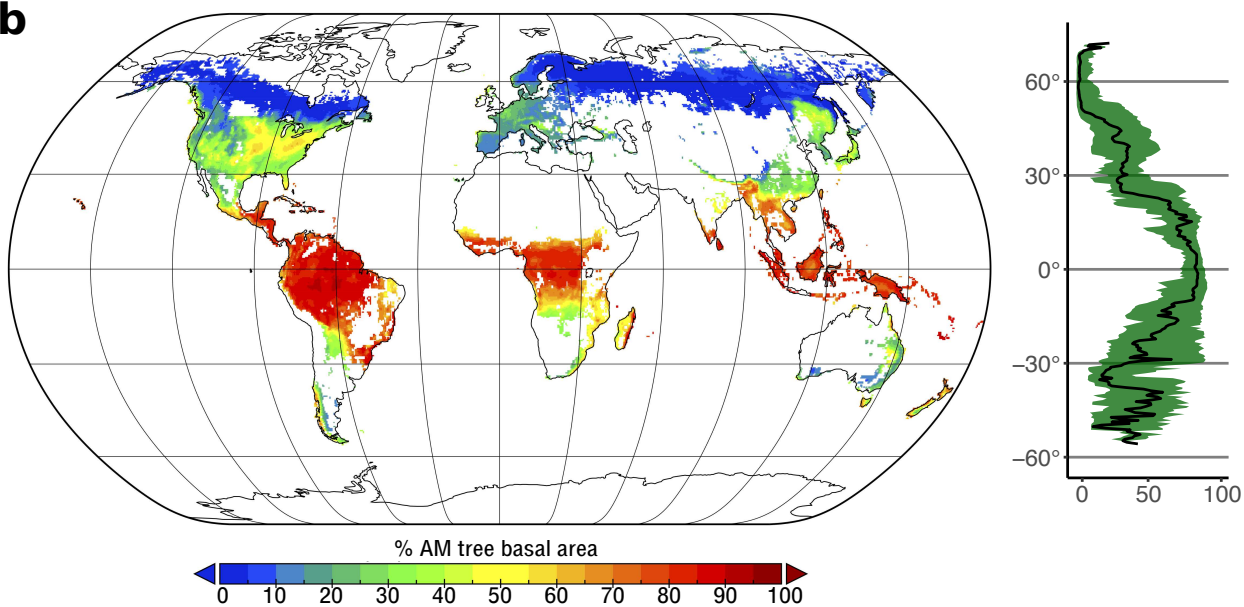
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